

CLERODANE DITERPENOIDS FROM *POLYALTHIA LONGIFOLIA**

ANIL P. PHADNIS, SARITA A. PATWARDHAN, NARAYANDATTA N. DHANESHWAR, SUDAM S. TAVALE and TAYUR N. GURU ROW

National Chemical Laboratory, Pune 411 008, India

(Revised received 19 January 1988)

Key Word Index—*Polyalthia longifolia*; Annonaceae; clerodane diterpenes; antifeedant activity.

Abstract—Two clerodane type diterpenoids, with antifeedant properties have been isolated from *Polyalthia longifolia* and identified as 16 α -hydroxy-cleroda-3,13(14)*Z*-dien-15,16-olide and 16-oxo-cleroda-3,13(14)*E*-dien-15-oic acid on the basis of spectral properties. Configuration of the olide at C-16 was established by X-ray crystallographic analysis.

INTRODUCTION

Polyalthia longifolia Thw (Annonaceae) is a tall, handsome, evergreen tree cultivated in gardens all over India. Earlier work on *P. longifolia* reports isolation of proanthocyanidin trimer [1] and sitosterol [2] from the bark. In continuation of our work on screening of plant species for pest control activity we found antifeedant activity in the acetone extract of leaves of *P. longifolia*. This note reports isolation and identification of two clerodane diterpenes **1** and **3**. X-ray analysis of the acetate (**2**) of **1** confirmed the stereochemistry at C-16.

RESULTS AND DISCUSSION

Air-dried leaves of *P. longifolia* were powdered and extracted with acetone. Chromatographic separation of the extract gave compounds **1** and **3**. Compound **1** (0.5%), C₂₀H₃₀O₃, exhibited four methyl signals; two tertiary, one secondary and one olefinic in the ¹H NMR spectrum (Table 1). The presence of hydroxy group (ν_{\max} 3345 cm⁻¹) and β -substituted butenolide [ν_{\max} 1730, 1635 cm⁻¹; δ H 5.83 (1H, s); δ C 171 (s), 117 (d) and 172 (s) (Table 2)] was evident from spectral data and suggested a

clerodane skeleton for **1**. A shift of a one proton singlet at δ 6.04 in the spectrum of **1** to 6.84 on acetylation and the presence of doublets at δ 99.69 and 94.04 in the ¹³C NMR spectra of **1** and **2**, respectively, indicated the presence of a hydroxy group at the C-16 position. Compound **1**, therefore, was identified as 16-hydroxy-cleroda-3,13(14)*Z*-dien-15,16-olide. Single X-ray diffraction analysis of the crystalline acetate derivative (**2**) established an α -configuration for the hydroxyl at C-16. Finally, compound **1** was designated as 16 α -hydroxy-3,13(14)*Z*-dien-15,16-olide.

The structure was solved by single crystal X-ray diffraction studies. Crystal data: crystals are monoclinic, space group *P*2₁, *Z* = 2, with *a* = 8.963 (2), *b* = 7.515 (2), *c* = 15.323 (1) Å, β = 97.38 (1)°. The data were collected with a 4-circle automatic diffractometer using MoK α (λ = 0.7107 Å). From 1651 independent reflections 1075 [*I* > 3 σ (*I*)] were considered as observed and used for structure solution. The structure was solved by direct methods [3] and refined by full matrix least squares refinement [4] in two blocks (anisotropic temperature factors for non-hydrogen atoms) with hydrogens in calculated positions which were confirmed by difference Fourier map. The isotropic temperature factors for hydrogen atoms were fixed. The final *R* factor was 0.065 after applying weights. The cyclohexene ring (ring A) has a 'half chair' and the cyclohexane ring (ring B) has a

* NCL Communication No. 4243.

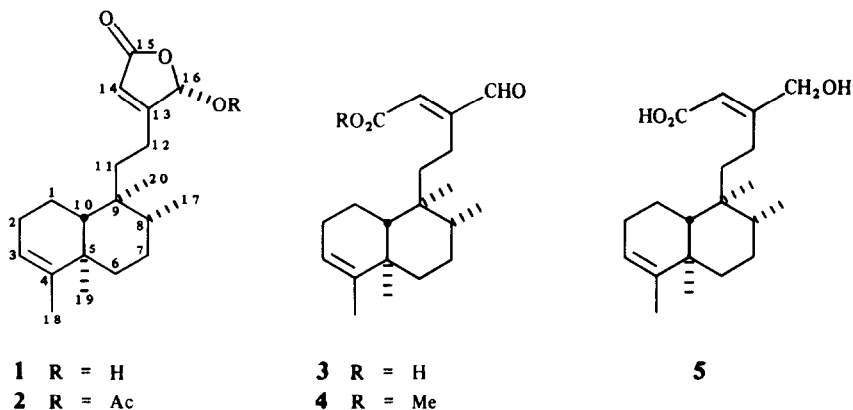


Table 2. ^{13}C NMR shifts (δ ppm) of compounds **1**, **2** and **4**

C	1	2	4	C	1	2	4
1	18.06	17.93	17.93	11	27.62 ^b	26.90 ^b	27.47 ^b
2	21.57	20.60	18.97	12	35.09	35.23	36.75
3	120.68	120.42	120.61	13	171	169.16	144.06
4	114.53	144.50	155.45	14	117.04	118.28	134.37
5	38.40 ^a	38.34 ^a	38.08 ^a	15	172	164.99	165
6	26.97 ^b	27.49 ^b	26.67 ^b	16	99.69	94.04	194
7	36.58	36.84	36.75	17	16.11	15.98	15.74
8	36.97	36.85	36.89	18	18.52	18.45	18.06
9	38.92 ^a	38.86 ^a	39.17 ^a	19	18.20	18.13	17.83
10	46.79	46.72	46.41	20	20.08	20.01	19.79
						OCOMe	-OMe
						21.25	51.75
						OCOMe	
						168.19	

^{a, b} Values in any vertical column may be interchangedTable 1. ^1H NMR spectral data (δ H ppm) of compounds **1–4**

H	1	2	3	4
2	2.01 <i>br</i>	2.08 <i>br</i>	2.06 <i>br</i>	2.1 <i>br</i>
3	5.2 <i>br</i>	5.17 <i>br</i>	5.15 <i>br</i>	5.2 <i>br</i>
12	2.24 <i>m</i>	2.17 <i>m</i>	2.53 <i>t</i>	2.5 <i>m</i>
14	5.83 <i>s</i>	5.73 <i>s</i>	6.37 <i>s</i>	6.4 <i>s</i>
16	6.04 <i>s</i>	6.84 <i>s</i>	9.46 <i>s</i>	9.5 <i>s</i>
17	0.83 <i>d</i> *	0.8 <i>d</i> *	0.73 <i>d</i> *	0.85 <i>d</i> *
18	1.61 <i>d</i> †	1.57 <i>d</i> †	1.5 <i>d</i> †	1.58 <i>d</i> †
19	1.0	1.0 <i>s</i>	0.93 <i>s</i>	0.97 <i>s</i>
20	0.75 <i>s</i>	0.75 <i>s</i>	0.62 <i>s</i>	0.66 <i>s</i>
-OMe	—	—	—	3.8 <i>s</i>
-OAc	—	2.17 <i>s</i>	—	—

* $J = 7$ Hz.† $J = 2$ Hz.

'chair' conformation. Since the torsion angles along the common C(5)–C(10) bond have opposite signs the two rings are *trans*-fused [5, 6]. A perspective view of the molecule is shown in Fig. 1. Full crystal data are deposited at the Cambridge Crystallographic Data Centre, University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, U.K.

A literature survey revealed that Bohlmann and co-workers have recently reported 16,16-dihydroxylkolava-3,13(14)*Z*-dien-15-oic lactone with ambiguous configuration at C-16 from *Acritopappus longifolius* [7].

Compound **3** (1%), $\text{C}_{20}\text{H}_{30}\text{O}_3$, showed four methyl signals [δ 0.62 (*s*), 0.93 (*s*), 0.73 (*d*) and 1.5 (*d*)] in its ^1H NMR spectrum similar to those in the spectrum of **1**. In the lower field region the 16-H singlet at δ 6.04 in the spectrum of **1** was replaced by a singlet at δ 9.46 in that of **3** and in the ^{13}C NMR spectrum of **1**, the 16-C doublet at δ 99.69 was shifted to δ 194 in the spectrum of **3** indicating replacement of hydroxyl by an aldehyde. The presence of

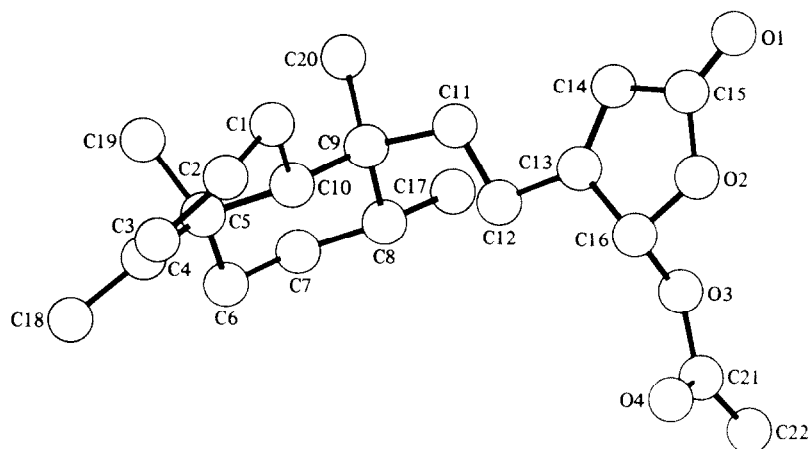
Perspective view of the molecule of compound **1**

Fig. 1.

α,β -unsaturated ester carbonyl as well as α,β -unsaturated aldehyde is revealed by peaks at ν_{\max} 1740, 1710 and 1650 cm^{-1} in the IR spectrum of methyl ester **4**. Sodium borohydride reduction of **3** afforded a primary alcohol, 16-hydroxy cleroda-3,13(14)*E*-dien-15-oic acid (**5**), which failed to lactonize on treatment with tosic acid, confirming the *E*-configuration of the C-13(14) double bond and it was found to be identical with the compound reported by Bohlmann [3]. Compound **3**, therefore, is identified as 16-oxocleroda-3,13(14)*E*-dien-15-oic acid.

Compounds **1** and **3** exhibited antifeedant activity against caterlooper (*Achaea janata*).

EXPERIMENTAL

^1H and ^{13}C NMR: at 90 MHz in CDCl_3 and TMS as internal standard, chemical shifts in ppm (δ). MS: 70 eV, direct inlet system.

Isolation of compounds 1 and 3. Green leaves of *P. longifolia* were dried in the shade and powdered. The powdered material (1 kg) was extracted with Me_2CO ($2 \times 5\text{ l}$) at room temp. The solvent was removed at $40^\circ/30\text{ mm Hg}$ in a rotavapour to yield a dark green extract (100 g). The extract (50 g) was treated with petrol ($2 \times 250\text{ ml}$) and the soluble portion (24 g) was chromatographed over silica gel. The column was successively eluted with petrol-EtOAc (9:1) [fraction A1 (5.6 g)], petrol-EtOAc (4:1) [fraction A2 (7 g) in earlier fractions and fraction A3 (6.65 g) in later fractions] and Me_2CO [fraction A4 (4.35 g)] Fr. A2 was extracted with NaHCO_3 soln which on treatment with conc. HCl gave compound **3**. After passing through activated charcoal compound **3** was obtained as a colourless gum (5 g 1%) $[\alpha]_D^{26} -70.58^\circ$ (MeOH; c 0.0107). UV $\lambda_{\max}^{\text{MeOH}}$ 238 nm (ϵ 12935). IR $\nu_{\max}^{\text{Neat}}\text{ cm}^{-1}$: 3345, 2920, 1730, 1635, 1435, 1125, 940. MS m/z (rel. int.): 318 $[\text{M}]^+$ (10), 303 (6), 285 (15), 190 (100), 189 (19), 175 (25), 161 (20), 135 (48). Treatment of **3** with CH_2N_2 yielded methyl ester **4** as a colourless gum $[\alpha]_D^{26} -90.9^\circ$ (MeOH; c 0.088). UV $\lambda_{\max}^{\text{MeOH}}$ 238 nm (ϵ 9304). IR $\nu_{\max}^{\text{Neat}}\text{ cm}^{-1}$: 2990, 1740, 1710, 1650, 1450, 1390, 1020, 910, 880. MS m/z (rel. int.): 332 $[\text{M}]^+$ (10), 317 (8), 301 (12), 285 (8), 203 (14), 191 (78), 190 (100), 189 (90), 175 (32), 135 (49), 121 (65), 107 (68), 95 (98). (Found: C, 75.58; H, 9.50 $\text{C}_{21}\text{H}_{32}\text{O}_3$ requires C, 75.86; H, 9.70%.)

Fr. A3 on repeated chromatography over silica gel afforded **1** as a colourless gum (2.5 g, 0.5%). $[\alpha]_D^{26} -70.58^\circ$ (MeOH, c 0.0107). UV $\lambda_{\max}^{\text{MeOH}}$ 210 nm (ϵ 14163). IR $\nu_{\max}^{\text{Neat}}\text{ cm}^{-1}$: 3345, 2920, 1730, 1635, 1435, 1125, 940, 750. MS m/z (rel. int.): 318 $[\text{M}]^+$ (8), 303 (5), 285 (12), 191 (40), 190 (100), 189 (15), 175 (22), 135 (48), 123 (60), 107 (88), 94 (99).

Acetylation of compound 1. Compound **1** (1.2 g) in pyridine (10 ml) was treated with AC_2O (10 ml) and left at room temp. for

48 hr. The reaction mixture was poured into cold H_2O and extracted with EtOAc. The organic layer after washing with H_2O and brine was dried (Na_2SO_4) and evapd to give a semi-solid residue which was purified by chromatography over silica gel to give compound **2**, mp 175° (petrol + EtOAc) (0.92 g, 71%). $[\alpha]_D^{26} -24.24^\circ$ (MeOH; c 0.066). IR $\nu_{\max}^{\text{Neat}}\text{ cm}^{-1}$: 2900, 1760, 1640, 1450, 1380, 1200, 880, 725. MS m/z (rel. int.): 360 $[\text{M}]^+$ (1) 300 (27), 285 (88), 267 (13), 190 (40), 189 (100), 175 (40), 119 (40), 107 (61), 105 (58), 91 (53). (Found: C, 73.03; H, 9.12. $\text{C}_{22}\text{H}_{32}\text{O}_4$ requires C, 73.30; H, 8.95%.)

Reduction compound 3. A soln of **3** (0.2 g) in EtOH (8 ml) was treated with NaBH_4 (0.08 g) at 5° and left overnight. The reaction mixture was diluted with cold H_2O and extracted with EtOAc. The organic layer was washed with brine and dried (Na_2SO_4). After removing solvent the residue was chromatographed to give **5** (0.1 g), mp 170° (petrol + C_6H_6). $[\alpha]_D^{26} -67.85^\circ$ (MeOH; c 0.112). IR $\nu_{\max}^{\text{Neat}}\text{ cm}^{-1}$: 3280, 2950, 1695, 1640, 1450, 1380, 1290, 900. ^1H NMR: δ 0.75 (3H, s), 0.84 (3H, d, $J=7\text{ Hz}$), 1.03 (3H, s), 1.59 (3H, d, $J=2\text{ Hz}$), 2.06 (2H, br), 2.37 (2H, m), 4.18 (2H, d, $J=2\text{ Hz}$), 5.15 (1H, br), 5.96 (1H, s). MS m/z (rel. int.): 320 $[\text{M}]^+$ (2), 305 (4), 191 (42), 175 (11), 163 (12), 149 (21), 135 (32), 121 (55), 107 (100), 95 (95). (Found: C, 75.21; H, 10.18; $\text{C}_{20}\text{H}_{32}\text{O}_3$ requires C, 74.96; H, 10.06%.)

Acknowledgement—The authors are grateful to Prof. F. Bohlmann for the ^1H NMR (270 MHz) spectrum of compound **5**.

REFERENCES

1. Agrawal, S. and Misra, K. (1979) *Curr. Sci.* **48**, 141.
2. Manzoor-i-Khuda, M., and Hossain, M. M. Golam, (1982) *Bangladesh J. Sci. Ind. Res.* **17**, 134.
3. Main, P., Hull, S. E., Lessinger, L., Germain, G., Declercq, J. P. and Woolfson, M. M. (1978). MULTAN-78, A system of computer programs for the automatic solution of crystal structure from X-ray diffraction data. University of York Louvain.
4. Ganztzel, P. K., Sparks, R. A. and Trueblood, K. N. (1961), LALS Full matrix least squares refinements of positional and thermal parameters and scale factors University of California, Los Angeles.
5. Mukherjee, M. and Mukherjee, A. K. (1984), *Acta Crystallogr.* **C40**, 983.
6. Bucourt, H. (1974) in *Topic in Stereochemistry* (Elliel, E. L. and Allinger, N. L., eds) Vol. 8, pp. 224. Wiley, New York.
7. Bohlman, F., Jakupovic, A. J., King, R. M. and Robinson, H. (1984), *Rev. Latinoam. Quim.* **15**, 16.